T-CELL-, MHC I-, AND TUMOR-RELATED GENE SIGNATURES PREDICT CLINICAL BENEFIT AND RESISTANCE TO TISLELIZUMAB MONOTHERAPY IN PRETREATED PD-L1-HIGH UROTHELIAL CARCINOMA

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BACKGROUND

- Urothelial carcinoma (UC) is the most common histologic type of bladder cancer and has a low 5-year survival rate when diagnosed at an advanced stage 1
- In China, UC is the sixth most common cancer in men and the thirteenth most common in women²
- While several FDA-approved anti-programmed cell death protein-1/programmed death-ligand 1 (PD-1/PD-L1) therapies have exhibited improved efficacy in lateline UC,³ the mechanisms of response or resistance are not clear and warrant further exploration
- Tislelizumab, a humanized IgG4 monoclonal antibody with high affinity and binding specificity for PD-1⁴ is the first PD-1/PD-L1 blockade approved in PD-L1–high UC⁵ by the National Medical Products Administration (NMPA) in China (April 2020)
- Here, retrospective analyses of gene expression profiles correlating with response and resistance to tislelizumab treatment are presented

METHODS

Study Design

- This phase 2 trial (BGB-A317-204; NCT04004221) conducted in China and Korea assessed the safety, tolerability, and efficacy of tislelizumab (200 mg every 3 weeks) in patients with PD-L1–high UC previously treated with \geq 1 platinumcontaining therapy
- Patients were considered PD-L1-high if $\geq 25\%$ of tumor/immune cells expressed PD-L1 as assessed by the VENTANA[™] PD-L1 (SP263) assay
- One hundred Chinese patients had evaluable gene expression profiling (GEP) data and were further analyzed

Gene Expression Profiling

- GEP analysis was conducted on baseline tumor samples (formalin-fixed, paraffin-embedded) using HTG EdgeSeq Precision Immuno-oncology panel
- Gene signature scores were calculated using the Gene Set Variation Analysis package and publicly available gene signatures
- Non-responder subgroups were hierarchically clustered by Euclidean distance metrics with average linkage by columns

Statistical Analysis

- Objective response rate (ORR) and Clopper-Pearson 95% confidence intervals (Cls) were calculated
- The Kaplan-Meier method was used to estimate median overall survival (OS), progression-free survival (PFS), and 95% CIs
- Differential gene expression (DEG) analysis was performed between responders and non-responders using the Wilcoxon rank-sum test; the Bonferroni method was used to adjust for multiple comparisons
- Kaplan-Meier curves of biomarker subgroups were compared using the log-rank test; hazard ratios were estimated using the Cox proportional hazard model

RESULTS

Patient Characteristics

• Baseline characteristics and clinical outcomes of the 100 GEP-evaluable patients were comparable with the overall population in this trial (Table 1)

T-cell and MHC I Signatures Were Associated With Response and **Survival Benefits**

- T-cell⁶ (CD3D, CD3E, CD3G, CD6, SH2D1A, TRAT1) and major histocompatibility complex class I (MHC I) signatures⁷ (HLA.A, TAP1) were enriched in responders (Figure 1A and 1B)
- Using median signature score as the cutoff:
- Significant improvement of OS (median OS: 13.5 vs 7.3 months, P=0.013, Figure 1E) and a trend of longer PFS (median PFS: 3.8 vs 2.1 months, P=0.125, Figure 1C) was observed in the T-cell-high group versus the T-cell-low group
- Significant improvement of PFS (median PFS: 4.1 vs 2.1 months, P=0.016, Figure 1D) and a trend of longer OS (median OS: 11.4 vs 8.5 months, P=0.184, Figure 1F) was observed in the MHC I-high group versus the MHC I-low group

Table 1: Baseline

Characteristic

Age, n (%)

Sex, n (%)

Known metastasi n (%)

Number of previous regimens, n (%)

Response, n (%)

ORR, % (95% CI)^a Median PFS, mont

Median OS, mont

RECIST v1.1 in the overall /GEP-evaluable population. PR, partial response; SD, stable disease.



Characteristics and Clinical Outcomes			
	Overall (n=113)	GEP Evaluable (n=100)	
<65	44 (38.9)	34 (34.0)	
≥65	69 (61.1)	66 (66.0)	
Male	84 (74.3)	75 (75.0)	
Female	29 (25.7)	25 (25.0)	
Lymph node only	25 (22.3)	23 (23.0)	
^s , Liver metastasis	33 (29.5)	29 (29.0)	
Visceral metastasis	87 (77.7)	77 (77.0)	
1	69 (61.1)	61 (61.0)	
2	37 (32.7)	32 (32.0)	
≥3	7 (6.2)	7 (7.0)	
CR	10 (8.8)	10 (10.0)	
PR	15 (13.2)	13 (13.0)	
SD	15 (13.2)	12 (12.0)	
PD	51 (45.1)	47 (47.0)	
Non-CR/Non-PD	4 (3.5)	3 (3.0)	
NE	18 (15.9)	15 (15.0)	
A	22 (14.9, 30.9)	23 (15.2, 32.5)	
nths (95% CI)	2.2 (2.1, 3.9)	2.1 (2.1, 4.0)	
ths (95% CI)	9.8 (7.5, 12.5)	9.8 (7.5, 12.2)	

^aObjective response rate was the proportion of patients with confirmed complete/partial responses using

Abbreviations: CI, confidence interval; CR, complete response; GEP, gene expression profile; NE, not estimable; ORR, objective response rate; OS, overall survival; PD, progressive disease; PFS, progression-free survival;

the Longest Survival Benefit

- Correlation of T-cell and MHC I signatures were low (Figure 2A) • Using the median as a cutoff, three subgroups were identified by combining both signatures: double high (30/100), single high (40/100), and double low (30/100) • The double-high subgroup (T-cell-high and MHC I-high groups; ORR=40%) had higher ORR compared with either the single-high (ORR=20%, P=0.03) or
- double-low (ORR=10%, P=0.01) subgroups (Figure 2B)
- Significantly longer PFS (P=0.04) and OS (P=0.05) were observed for patients with double-high signature score (Figure 2C and 2D)



comparisons between subgroups for PFS (C) and OS (D) by utilizing the double high group as the reference. Abbreviations: BEP, biomarker evaluable population; CI, confidence interval; HR, hazard ratio; MHC I, major histocompatibility complex class I; mOS, median overall survival; mPFS, median progression-free survival; NE, not estimable; OR, odds ratio; ORR, objective response rate; OS, overall survival.

Double-High Signature Score Subgroup Showed the Highest ORR and

Differential Signature and Single Gene Expression Between Responders and Non-responders

- NR3) according to immune- and tumor-related gene signatures (Table 2) significantly higher expression of T-cell co-inhibition signatures (Figure 3A, P=0.04)
- Non-responders were clustered into three distinct subgroups (NR1, NR2, and • With a comparable tumor inflammation signature (TIS) to responders, NR1 had
- NR2 was enriched for TGF β signaling (Figure 3C, P=0.004)
- NR3 exhibited non-inflamed features (decreased TIS [Supplemental Figure 1, P=0.01]), but was enriched for cell cycle signatures (**Figure 3E**, P=0.0003) • Key genes enriched in each non-responder group are shown in Figure 3B, 3D, and **3**F

Poster: 78 Society for Immunotherapy of Cancer November 9-14, 2020, Virtual Congress

CONCLUSIONS

- T-cell and MHC I signatures were associated with better clinical response and longer PFS and OS after tislelizumab treatment
- Combining high T-cell signatures with high MHC I signatures identified a subgroup with improved clinical benefit (40% ORR, 5.26-month median PFS and 15.2-month median OS)
- Distinct gene expression profiles were negatively correlated with clinical benefit, indicating the presence of a potential resistance signal:
- Elevated immune checkpoint and suppressive immune cell signatures in inflamed non-responders
- High expression of TGF β signaling and angiogenesis-related genes
- Low tumor inflammation signature with enriched cell cycle signature • The association between gene expression biomarkers and clinical benefit could enhance our understanding of the process of adaptive antitumor
- immunity and also indicate a potential novel combination treatment strategy • Due to the limitations associated with a single-arm study, the potential predictive role of the gene signatures discussed above warrant further exploration in an ongoing phase 3 study (RATIONALE 310; BGB-A317-310; NCT03967977)

Non-responders Could Be Clustered Into Three Distinct Subgroups According to the Immune- and Tumor-related Gene Signatures Table 2: Immune and Tumor Gene Signatures Utilized for Non-responder Subgroups Clustering

Antitumor Activity	Immune Cells		Feature of Tumor
TIS	Macrophage	CD8 T cell	Endothelial cells
Cytotoxic activity	M1 Macrophage	T cell	Focal adhesion
IFNγ	M2 Macrophage	DC	EMT
MHC class I	Mast cell	NK	Angiogenesis
Checkpoint-T cell	Neutrophils	MDSC	TGF β signaling
Checkpoint-APC		B cell	Cell cycle

Abbreviations: APC, antigen presentation cell; DC, dendritic cell; EMT, epithelial-mesenchymal transition, MDSC, myeloid-derived suppressor cell; MCH, major histocompatibility complex; NK, natural killer cell; TIS, tumor inflammation signature.

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ACKNOWLEDGMENTS

The authors wish to acknowledge the investigative centers' study staff and study patients, and to recognize those from BeiGene, Ltd. who have substantially contributed to the development of this presentation. This study was sponsored by BeiGene,

Ltd. Editorial assistance was provided by Stephan Lindsey, PhD, and Elizabeth Hermans, PhD (OPEN Health Medical Communications) Chicago, IL), and funded by the study sponsor.

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